

Original Research Article

Nuclear anomalies in the buccal cells of children under dental treatment

Elisângela de Fátima Rezende¹ Maria Cristina Mendes-Costa¹ Johnson Campideli Fonseca¹ Alex de Oliveira Ribeiro¹

Corresponding author:

Maria Cristina Mendes-Costa University Center of Lavras 506, Padre José Poggel St. – Centenario Zip code 37200-000 – Lavras – MG – Brazil E-mail: mcmcosta@unilavras.edu.br

¹ Center of Researches in Health and Biological Science, University Center of Lavras – Lavras – MG – Brazil.

Received for publication: August 17, 2010. Accepted for publication: November 3, 2010.

Keywords: genotoxicity; micronucleus tests; dental materials.

Abstract

Introduction: Since some dental materials may be aggressive to a person's body, studies involving such materials seem to be necessary. **Objective:** This study was conducted to evaluate the genotoxicity of dental materials through micronucleus (MN) test. Material and methods: Exfoliated buccal cells of 4-to-12 year-old children, who were on some type of dental treatment, were collected either before or after the treatment ending. Each sample was composed of 1,000 cells per patient. Student's t test was used for comparison. Results: The dental materials were divided into 3 groups, as follows: cement, monomers, and their combination. Treatments using monomer + cement-based materials were found to increase significantly the number of binucleated (BN) cells, (p < 0.05) which indicate several degenerative nuclear changes. Conclusion: The combination of cement-based dental material with monomers increases the cytotoxic action of dental materials.

Introduction

Dental materials may produce aggressive effects caused by monomers release and/or other organic and inorganic components [31]. As most dental materials release small portions of several substances on both the pulp and oral cavity, it is essential to prove their biocompatibility and toxicological profile. Appropriate regulation should ensure that the genotoxicity be either abolished or decreased [17].

The main dental materials used in pediatric dental treatment are: glass ionomer cements (GIC), zinc oxide - eugenol (ZOE), and resin composites (RC). GIC maintains the surrounding environment suitable for remineralization as well as interferes with the bacteria metabolism by fluoride releasing [2, 26]. They are employed as pulp capping, occlusal sealant, and restorative material. Also, hydrophilic monomers can be incorporated to GIC, so-called resin-modified glass ionomers (RMGICs). The polymerization reactions make monomers into polymers. Consequently, the residual monomers released may cause intense cytotoxic effects [9, 34]. ZOE is often used in low-cost, easy-dealing temporary restoration [7]. Its effects can vary, as follows: point mutation inducer [27]; cytotoxicity [18, 27]; antimicrobial [6, 31]. RC is commonly used in permanent restoration. It presents less abrasion, offering easy handling. It also can be found in different colors to match to the tooth to be restored [3]. These materials may have some genotoxic potential. An increasing number of dental restorations have been performed over the past decade with significantly increase in local and systemic adverse effects such as cytogenetic changes [29].

Chromosome damages are widely used as biomarkers in monitoring human exposure to carcinogenic agents [8, 20, 36]. Binucleated (BN) cells may indicate several kinds of degenerative nuclear changes [10]. BN are cells which have two similarsized nuclei (almost the same size). Such nuclei are not overplaced, but may be side by side. They have the same color, an intact nuclear membrane, and are within the cytoplasm [14]. Micronuclei (MN) are free round or egg-shaped corpuscles, about 1/3 to 1/16 of the nucleus size [4, 5, 25]. They are usually found beside the main nucleus and are similar in shape, color, and chromatinic body distribution [14]. Such structures are a result of chromosome fragments or entire acentric chromosomes which are lost during a cell division. For this reason they are not included in the daughter cells' nuclei, thus remaining in the cytoplasm of interphase cells [5, 10, 16, 35]. MN takes 7-16 or 30 days to be formed [6, 28, 33, 37, 38]. Such period is related to the amount of time that the basal cells take to reach the surface and exfoliate.

It is extremely important to know how the materials act in human tissues because they are in close contact with oral mucosa [30]. This study aims to evaluate the genotoxicity of dental materials used in pediatric treatment through micronucleus (MN) test of exfoliated buccal cells.

Material and methods

Study Design

The study was approved by the Ethics Committee of Unilavras (CAAE, 0036/06). The sample comprised 72 children divided into control group (n = 40) and test group (n = 32). Inclusion criteria during this study were as follows: children who had never undergone any dental treatment; children aged 2-12 years (mean age of five years). There were no children with neurological diseases and related genetic alterations. Their participation in the research was previously authorized by their parents or children's legal guardians.

Exclusion criteria comprised children with damages in oral mucosa that preclude the collection of cells and whose parents had not signed the clarified consent form or did not want to participate in the study.

Test group patients were divided into 3 experimental subgroups according to the dental material used, as follows: Group I - Cement-type materials: glass ionomer cement and zinc oxide cement + eugenol (n = 8); Group II - Monomerbased materials: composite resin, dental sealant, and adhesives (n = 13); Group III: combination of both (n = 11). The genetic material used was obtained by collecting oral mucosa cells by scrubbing the surface with a wood stick. The first collection was done before the treatment in order to find out the basal index of both micronucleus and binucleus cells likely to be found in the mucosa.

One month after the treatment, a new sample collection was performed in the treated patients when they returned for either a follow-up or new treatment appointment This time period was chosen because is related to the mitoses that the altered cells would suffer, thus showing clearly their micronucleus formation. The control group consisted of patients' oral mucosa cells before the insertion of any type of dental material.

Genotoxicity evaluation (micronucleus test)

After the inner part of the cheek was scrubbed by using a wood stick [10, 38], the epithelial cells collected from buccal mucosa were smeared onto clean microscope glass slides fixed in 70% alcohol, air dried, and stained with 2% acetic orcein [15], for 30' [23]. A light microscope at 100 X magnification on coded slides was used for MN analysis and then microphotographed (Samsung SDC-312). Two thousand cells per patient were analyzed, as follows: 1,000 before and 1,000 after the treatment [19, 24]. After both the micronucleus and the binucleated cells were quantified, the frequency analysis of both alterations was performed only in the non fragmented, overplaced or overcrowded cells with untouched nuclei, according to the acceptance criteria, as described by Fenech et al. (2000). Data statistic analysis was performed by Student's t test [19] for non-parametric data.

Results and discussion

The epithelial tissue of the oral cavity was collected for the test, because it is in close contact with dental materials and is constantly renewed [1]. In addition to this, it can act as a tool for biomonitoring human populations exposed to genotoxic agents [12, 22].

The MN average frequency in a healthy population is about 1 to 3 out of 1,000 cells [12]. However, in our study, nearly 1% of the control patients exhibited MN score of 7 to 8 before treatment (figure 1). This suggests that those children had probably been influenced by genotoxic agents [1, 22, 31].

Averages between control and treated groups



Figure 1 - Mean values of BN and MN average in control and experimental groups

Normal, binucleated, and micronucleated cells were found in all studied groups. The binucleated cells (figure 2) presented two nuclei, nearly the same size instead of overplaced. They showed the same color, their nuclear membrane was intact and within the same cytoplasm, as described by Fenech *et al.* (2003). On the other hand, the nucleus of the micronucleated cells (figure 3) was shorter than the main nucleus diameter, round or oval in shape, separated from the main nucleus [14]. Diler and Ergene [11] analyzed nuclear anomalies in buccal cells of calcite factory workers and found micronucleus and binucleated cells suggesting significant cytogenetic damage.



Figure 2 - Binucleated cell



Figure 3 – Micronucleated cell

The cement materials did not caused decrease and/or increase of BN or MN cells in Group I patients (table I). The treatment with monomers materials (Group II) did not significantly alter the frequency of MN, because pre- and post-treatment MN frequency score was similar (table I).

It is important to consider that both MNs and BNs have a time-determined formation. The dental

material constantly releases small portions of its own substances in the oral cavity [17], especially during the first 30 days after its insertion [2]. Therefore, the time period of this study was enough for all the possible variations appear.

The increase of the frequency of BN for Group III was significantly influenced by the cement + monomer combination (table I). Similarly results were found in people working in oil and oil-products stations [11] and in between smoking and exposure to benzene workers [12].

Many materials, particularly root canal sealers, remain in contact with vital tissues for a long period, when cellular aggression by chemical, physical and mechanical elements may occur, being the respiratory system one of the first cells to be affected. Some aggressor agents, particularly chemical substances, block important enzyme systems of the protein synthesis and/or the generation of ATP; others lead to the generation of harmful intracellular products or, still, act directly destroying vital structural components of the cell [30]. This may justify the appearance of binucleated cells (Group III). Senne et al. (2009) analyzed zinc oxide-eugenol cements and resin, finding that all tested sealers were cytotoxic. Reis et al. [24] studied the genotoxic effect of ethanol on oral mucosa cells and observed that the frequency means of micronucleated cells and micronuclei were significantly higher in the group of exposed individuals, when compared to the control group.

Group	BN		MN	
	Mean \pm SE	t Test 5%	Mean ± SE	t Test 5%
I – Treatment	2.180 ± 0.714	0.0922 (NS)	1.330 ± 0.122	0.8928 (NS)
I – Control	1.355 ± 0.760		1.299 ± 0.356	
II – Treatment	2.019 ± 0.375	0.0531 (NS)	1.696 ± 0.674	0.7913 (NS)
II – Control	1.441 ± 0.541		1.790 ± 0.486	
III – Treatment	2.413 ± 0.644	0.0069 (S)	1.821 ± 0.504	0.9371 (NS)
III – Control	1.588 ± 0.177		1.791 ± 0.722	

Table I - Evaluation of BN and MN in Groups I, II and III

(NS) Not significant

(S) Significant. The treatment values were significantly different from the control at p < 0.05

(SE) Standard Error

Celik *et al.* (2003) and Fenech *et al.* (1999) state that binucleated cells indicate cytotoxicity. The resin-modified glass ionomer cements (RMGIC) have hydrophilic monomers incorporated to the glass ionomer cement. When RMGICs are subpolymerized, they convert monomers into polymers. In such case

the released residual monomers may produce an intense cytotoxic effect [34]. The RMGIC cytotoxic effect was noticed in MDPC-23 odontoblastic cells culture [9]. When combined, the chemical composition of cements + monomers becomes similar to that of RMGIC. For this reason, group III materials were found to be related to cytotoxic damages. Such results were reached by comparing the number of BNs found either before or after the exposure to dental material. Although BNs are not directly involved with DNA, they interfere with the late events occurring in cell division [11, 39]. Because their consequences are still unknown, further cytogenetic studies regarding to the perpetuation of such cells in the oral mucosa are necessary.

In order to analyze genotoxicity or cytotoxicity in lymphocyte culture, the use of cytochalasin B is needed. This substance paralyses the cytokinesis, which promotes the nucleus division, resulting in a two-nucleus cell [13, 41]. The cement + monomer combination used in this study showed significant result for BN, suggesting a direct influence on cytokinesis similarly to cytochalasin B. Its action can be explained by the inhibition of telophase, consequently the cell reaches the epithelial surfaces with two nuclei. Because the action of the evaluated substances interferes directly into the cell instead of the gene, a cytogenetic damage occurs [11].

Biological markers may express the dose amount of exposure to carcinogens and their interaction with macromolecules, as DNA [24]. Consequently, a greater emphasis should be given to methods that detect the genotoxic human activity. The biomarkers may be used for the prevention of serious diseases, and detection of high-risk patients.

Conclusion

It can be concluded that a significant BN increase was noticed when Group III materials were used, suggesting that the combination of a cement-based dental material with monomers intensify the cytotoxic action. The combination of MN analysis with other nuclear anomalies, such as BN, was found to enhance the sensitiveness as well as the evaluation of both cytotoxicity and genotoxicity in scrubbed cells of the exfoliated buccal mucosa.

References

1. Arlett CF, Ashby J, Fielder RJ, Scott D. Micronuclei origins, applications and methodologies – a workshop sponsored by the Heath and Sfety Executive held in Manchester. Mutagenesi. 1989 May;4(6):482-5.

2. Bernardo PC, Rodrigues CR, Souza Paiva JA, Singer TM, Sañudo A. Avaliação clínica de um cimento de ionômero de vidro utilizado como selante oclusal. Pes Odontol Bras. 2000 Jan-Mar;14(1):53-7.

3. Bianchi EC, Aguiar PR, Poggi MR, Salgado MH, Freitas CA, Bianchi ARR. Estudo do desgaste abrasivo das resinas compostas disponíveis no mercado brasileiro. Mat Res. 2003 Apr-Jun;6(2):255-64.

4. Bindu L, Balarm P, Mathew A, Remani P, Bhattathiri VN, Nair MK. Radiation-induced changes in oral carcinoma cells – a multiparametric evaluation. Cytopathol. 2003 Oct;14(5):287-93.

5. Blochingm M, Hofmann A, Lautenschläger CH, Berghaus A, Grummt T. Exfoliative cytology of normal buccal mucosa to predict the relative risk of cancer in the upper aerodigestive tract using the MN-assay. Oral Oncol. 2000 Nov;36(6):550-5.

6. Carrano AV, Natarajan AT. Considerations for population monitoring using cytogenetics techniques ICPEMC publication 14. Mutat Res. 1988;204:379-406.

7. Cassanho ACA, Fernandes AM, Oliveira LD, Carvalho CAT, Jorge AOC, Koga-Ito CY. In vitro activity of zinc oxide-eugenol and glass ionomer cements on Candida albicans. Braz Oral Res. 2005 Apr-Jun;19(2):134-8.

8. Cerqueira EM, Gomes-Filho IS, Trindade S, Lopes MA, Passos JS, Machado-Santelli GM. Genetic damage in exfoliated cells from oral mucosa of individuals exposed to X-rays during panoramic dental radiographs. Mutat Res. 2004 Aug;562(1-2):111-7.

9. Coimbra LR, Giro EMA, Aranha AMF, Costa CAS. Citotoxicidade de cimentos de ionômero de vidro restauradores sobre células de linhagem odontoblástica. Rev Odont Ciên. 2006;21(54):338-45. 10. Çelik A, Çavas T, Ergene-Gözükara S. Cytogenetic biomonitoring in petrol station attendants: micronucleus test in exfoliated buccal cells. Mutagenesis. 2003 Sep;18(5):417-21.

11. Diler SB, Ergene S. Nuclear anomalies in the buccal cells of calcites factory workers. Genet Mol Biol. 2010 Jun;33(2):374-8.

12. Fenech M, Holland N, Chang WP, Zeiger E, Bonassi S. The Human Micronucleus Project. An international collaborative study on the use of micronucleus technique for measuring DNA damage in humans. Mutat Res. 1999 Jul;428(1-2):271-83.

13. Fenech M. The in vitro micronucleous technique. Mutat Res. 2000 Nov;455(1-2);81-95.

14. Fenech M, Chang WP, Kirsch-Volders M, Holland N, Bonassi S, Zeiger E. HUMN project: detailed description of the scoring criteria for the cytokinesis-block micronucleus assay using isolated human lymphocyte cultures. Mutat Res. 2003 Jan;534(1-2):65-75.

15. Guerra M, Souza MJ. Como observar cromossomos: um guia de técnicas em citogenética vegetal, animal e humana. 1. ed. Ribeirão Preto: Funpec; 2002. 131 p.

16. Heddle JA, Cimino MC, Hayashi M, Romagna F, Shelby MD, Tucker JD et al. Micronuclei as an index of cytogenetic damage: past, present, and future. Environ Mol Mutagen. 1991 Aug;18(4):277-91.

17. Heil J, Reifferscheid G, Waldmann P, Leyhausen G, Geurtsen W. Genotoxicity of dental materials. Mutat Res. 1996 Jul;368(3-4):181-94.

18. Hume WR. Effect of eugenol on respiration and division in human pulp, mouse fibroblasts and liver cells in vitro. J Dent Res. 1984 Nov;63(11):1262-5.

19. Kehdy FSG, Cerqueira EMM, Bonjardim MB, Camelo RM, Castro MCL. Study of the cytogenetic effects of occupational exposure to pesticides on sanitation workers in Belo Horizonte, Brazil. Genet Mol Res. 2007 Sep;6(3):581-93.

20. Marcon F, Zijno A, Crebelli R, Carere A, Veidebaum T, Peltonen K et al. Chromosome damage and aneuploidy detected by interphase multicolour FISH in benzene-exposed shale oil workers. Mutat Res.1999 Sep;445(2):155-66. 21. Munerato MC, Sinigaglia M, Reguly ML, Andrade HHR. Genotoxic effects of eugenol, isoeugenol and safrole in the wing spot test of Drosophila melanogaster. Mutat Res. 2005 Apr;582(1-2):87-94.

22. Pastor S, Gutiérrez S, Creus A, Xamena N, Piperakis S, Marcos R. Cytogenetic analysis of Greek farmers using the micronucleous assay in peripheral lymphocytes and buccal cells. Mutagenesis. 2001 Nov;16(6):539-45.

23. Pires NM, Souza IRP, Prates HT, Faria TCL, Pereira Filho IA, Magalhães PC. Effect of leucaena aqueous extract on the development, mitotic index and peroxidase activity in maize seedlings. Rev Bras Fisiol Veget. 2001 Apr;13(1):55-65.

24. Reis SRA, Sadigursky M, Andrade MGS, Soares LP, Espírito Santo AR, Vilas Boas DS. Genotoxic effect of ethanol on oral mucosa cells. Pesq Odontol Bras. 2002 Jul-Sep;16(3):221-5.

25. Ribeiro LR, Salvatori MF, Marques EK. Mutagênese ambiental. Canoas: Ulbra; 2003. 356 p.

26. Sacramento PA, Papa AMC, Carvalho FG, Puppin-Rontani RM. Propriedades antibacterianas de materiais forradores – revisão de literatura. Rev Odontol Unesp. 2008;37(1):59-64.

27. Sarrami N, Pemberton MN, Thornhill MH, Theaker ED. Adverse reactions associated with the use of eugenol in dentistry. Braz Dent J. 2002 Sep;14(5):257-9.

28. Sarto F, Tomanin R, Giacomelli L, Canova A, Raimondi F, Ghiotto C et al. Evaluation of chromosomal aberrations in lymphocytes, oral mucosa and hair root cells of patients under antiblastic therapy. Mutat Res. 1990 Feb;228(2):157-69.

29. Schweikl H, Hiller KA, Bolaya C, Kreissl M, Kreismanna W, Nusser A et al. Cytotoxic and mutagenic effect of dental composite materials. Biomat. 2005 May;26:1713-9.

30. Senne MI, Lemos N, Fidel SR, Fidel RAS. Evaluation of the cytotoxicity of three root canal sealers used in obturation of radicular canals system. RSBO. 2009 Mar;6(1):71-6. 31. Silva J, Erdtmann B, Henriques JAP. Genética toxicológica. Porto Alegre: Alcance; 2003. 442 p.

32. Spahl W, Budzikiewicz H. Qualitative analysis of dental resin composites by gas and liquid chromatography/mass spectrometry. Fresenius J Analy Chem. 1994 Nov-Dec;350(12):684-91.

33. Squier CA, Finkelstein MW. Mucosa bucal. In: Ten Cate AR. Histologia bucal. Desenvolvimento, estrutura e função. 5. ed. Rio de Janeiro: Guanabara Koogan; 2001. p. 323-39.

34. Stanislawski L, Daniau X, Lauti A, Goldberg M. Factors responsible for pulp cell cytotoxicity induced by resin-modified glass ionomer cements. J Biomed Mater Res. 1999 May;49(3):277-88.

35. Stich HF, Curtis JR, Parida BB. Application of micronucleus test to exfoliated cells of high cancer risk groups: tabacco chewers. Int J Cancer. 1982 Nov;30(5):553-9.

36. Stich HF, Rosin MP. Quantitating the synergistic effect of smoking and alcohol consumption with the micronucleus test on human buccal mucosa cells. Int J Cancer. 1983 Mar; 31(3):305-8.

37. Stich HF, Rosin MP. Micronuclei in exfoliated cells as a tool for studies in cancer risk and cancer intervention. Cancer Lett. 1984 Apr;22(3):241-53.

38. Tolbert PE, Shy CM, Allen JW. Micronuclei and other nuclear anomalies in buccal smears: a field test in snuff users. American J Epidemiol. 1991 May;134(8):840-50.

39. Tolbert PE, Shy CM, Allen JW. Micronúcleo e outras anomalias no espaço oral: métodos e desenvolvimento. Mutat Res. 1992;271:69-77.

40. Vasconcelos KRF, Veiga Júnior VF, Rocha WC, Bandeira MFCL. Avaliação in vitro da atividade antibacteriana de um cimento odontológico à base de óleo-resina de Copaifera multijuga Hayne. Braz J Pharmacog. 2008 Dec;18(1):733-8.

41. Zalacain M, Sierrasesúnaga L, Datiño A. El ensayo de micronúcleos como medida de inestabilidad genética induzida por agentes genotóxicos. Anales. 2005 [cited 2007 Sep 30]; 28(2). Available from: URL:www.cfnavarra.es/ salud/anales/textos/vol28/n2/revis2a.html.